#### **ORIGINAL PAPER**



# **Fly family diversity shows evidence of livestock grazing pressure in Mongolia (Insecta: Diptera)**

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Received: 5 January 2018 / Accepted: 4 April 2018 © The Author(s) 2018

### **Abstract**

Members of the insect order Diptera respond differentially to environmental changes and may play an important role in understanding the effects that livestock grazing disturbances have on biodiversity. Here we examine how increasing grazing pressures on the Mongolian steppe affect Diptera diversity and abundance. Using 2334 yellow pan traps, we sampled a total of 132 sites over four years to collect 17,348 flies. We compared fly diversity and abundance at five levels of livestock grazing. We observed that fly family diversity decreased in heavily grazed sites and that diptera communities at sites with intense grazing have proportionally higher prevalence of taxa from the families Muscidae, Sepsidae, Ephydridae, Chloropidae, and Tachinidae, two of which are often associated with animal waste. Chironomidae, Ceratopogonidae, Sarcophagidae, and Sciaridae are most prevalent at sites with very little or no grazing, and Anthomyiidae, Calliphoridae, Carnidae, Cecidomyiidae, Dolichopodidae, Empididae, Scatopsidae and Sphaeroceridae are most often encountered at sites with intermediate amounts of grazing. Observing changes in a few guilds of fly families at different grazing levels is beneficial in understanding human effects on fly diversity.

**Keywords** Diptera · Livestock grazing pressure · Disturbance · Family diversity · Mongolian Aquatic Insect Survey

## **Introduction**

The diversity of Diptera, a major group in terrestrial and freshwater ecosystems, can tell us much about how organisms respond to environmental disturbance. Understanding how environmental disturbance events affect biological diversity remains an active area of research in community ecology (Moctezuma et al. [2016](#page-11-0); Solar et al. [2016\)](#page-11-1). One such anthropogenic impact on the environment, livestock

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grazing, can be particularly detrimental to biological communities, especially in concentrated amounts (Papanastasis et al. [2017](#page-11-2)), but recognizing the consequences of slight or moderate grazing on the biodiversity of a community can be challenging. Livestock grazing results in the removal of ground cover, the deposition of fecal matter, and the compaction of soil, which can severely degrade habitats especially in combination with watershed and stream channel erosion (Yadamsuren et al. [2015](#page-12-0)). This overuse of land often results in significant reductions in diversity and productivity in both plants and animals (Milton et al. [1994\)](#page-11-3), but can also lead to the opposite outcome in terms of biotic diversity. Increasing available resources, such as fecal matter, can increase arthropod diversity by adding new niches (Verdú et al. [2000\)](#page-12-1). However, converting a diversity of resources to this single "fecal" resource could decrease habitat heterogeneity and thus niches available to diverse arthropods. Because specialists exist at both high and low levels of grazing, the intermediate disturbance hypothesis suggests that, in fact, the greatest amount of diversity should be found at intermediate levels of disturbance (Connell [1978\)](#page-11-4). These different biotic responses make it difficult to gauge the effect that livestock grazing has on biodiversity as a whole. We

therefore examine the effect that grazing has on one particularly diverse group of invertebrates, the insect order Diptera (true flies).

Invertebrates are overwhelmingly dominant in abundance and niche diversity in most biotic communities and for this reason provide a good avenue for analyzing the effect of overgrazing on an ecosystem (Taylor and Doran [2001\)](#page-12-2). In some heavily grazed areas, invertebrate diversity is low, but the abundance of a few taxa is much higher than in lightly grazed areas (Seymour and Dean [1999](#page-12-3)), probably because many invertebrates, including flies, are saprophagous or coprophagous by nature. Many flies rely on excrement provided by livestock for nutrition and reproduction, and thereby increase in population with increased grazing (McAlpine et al. [1981\)](#page-11-5). The enhancement of the detritivore and decomposer resources and cycles could increase the number of exploiting taxa.

As some of the most abundant and diverse invertebrates, true flies have been used to monitor ecological pressures (Servia et al. [1998](#page-12-4); Haslett [2001;](#page-11-6) Cabrini et al. [2013\)](#page-11-7). Among adult aquatic insects found on Mongolian streambanks, Diptera were most significantly affected by livestock grazing and erosion resulting in a decrease in richness (Hayford and Gelhaus [2010\)](#page-11-8). Crane flies (Tipuloidea) in particular were negatively impacted by grazing in Mongolia because of decreased plant biomass and desiccation of moss, a common larval habitat (Yadamsuren et al. [2015](#page-12-0)). Soil-dwelling Diptera have also been used as effective bioindicators because of the role they play in decomposition, their widespread abundance, and their niche similarity to other soil-dwelling species (Frouz [1999\)](#page-11-9). Where manure is present, there is an increase in abundance of soil-dwelling larvae, especially in the families Chironomidae, Sciaridae, Scatopsidae and Psychodidae (Frouz [1999\)](#page-11-9). However, the effect that heavy grazing has on adult fly taxonomic families and family diversity is unknown.

Mongolia has an array of diverse and, as yet, lightly polluted environments. With its low population density and varied biogeographical landscape, it has retained much of the biota that has been lost in surrounding more populous Asian countries (Fernandez-Gimenez [2000\)](#page-11-10). Nomadic grazing has been prominent in Mongolia for centuries, including ruminants such as yaks, sheep, goats and cattle, and nonruminating horses and camels (Johnson et al. [2006](#page-11-11)). The recent increase in the price of cashmere and privatization of land has led to domestic grazing becoming commercialized with heavier grazing in more areas (Mearns [2004;](#page-11-12) Maasri and Gelhaus [2011](#page-11-13)). This recent transition lends an opportunity to observe the short-term effects of grazing in Mongolia (Maasri and Gelhaus [2011\)](#page-11-13).

We investigated the effect of grazing pressure on terrestrial adult fly diversity and abundance to test whether flies can be used as bioindicators to evaluate human impact on habitats in the Mongolian steppe and taiga. Because aquatic flies responded negatively to streambank erosion (Hayford and Gelhaus [2010](#page-11-8)) which is closely related to grazing, we hypothesize that terrestrial adults have a measured response to grazing pressures as well. This response could be negative—meaning that fly diversity decreases in response to increased grazing pressures, intermediate—meaning that fly diversity is highest in areas that are moderately grazed, or positive—indicating that fly diversity is greatest in areas that have been heavily grazed. These response categories can also be applied at the family level, comparing how frequently a family is encountered at the above grazing intensities. We asked which families of flies are most heavily affected by grazing and which are best used as bioindicators of grazing pressures and found a diversity of responses in the fly families examined.

## **Materials and methods**

### **Study area**

We sampled fly communities near streams and rivers from sites across northern Mongolia (Fig. [1](#page-2-0); Table [1](#page-3-0)) as part of a larger general aquatic ecology survey of more than 400 sites called the Mongolian Aquatic Insect Survey (MAIS). This is an effort led by a group of international and Mongolian scientists to inventory aquatic insect diversity in the mountainous regions of Mongolia. As we visited these sites we also quickly sampled the terrestrial community. Collecting sites for this study were distributed primarily in the Altai range of western Mongolia, the Hangai range of central Mongolia, and the Onon river drainage of eastern Mongolia, and were sampled in July for four consecutive years (2008–2011). We selected sites based largely on relative ease of access on loops of rough roads that included many habitat types.

At each site, we sampled insect communities and took measurements of habitat conditions and biochemistry. Grazing intensity at each site was scored from 0 to 4 using a standardized rubric based on vegetation height and the composition of the grass community (Table [2](#page-5-0)). This minimized sampling bias by different researchers, an imperative as team members varied over the course of this project.

We collected insects at each site using yellow pan traps, a simple, cost-effective method to sample riparian flies (Vrdoljak and Samways [2012](#page-12-5)). With this approach, many flying insects, including flower-visiting guilds of insects as well as adults of aquatic insects, were trapped in a low-effort, highyield manner. Yellow plastic 12 oz bowls (SOLO® Number PSB2Y-0099) were placed in a line of pans parallel to the riverbank at 10 m intervals, usually less than 1 m from the water, and filled with water and a dish soap surfactant. The number of pans placed at each site varied over the 4 years,



<span id="page-2-0"></span>Fig. 1 Localities in Mongolia where pan trap data was collected from 2008 to 2011. Source of maps: Esri, DigitalGlobe, GeoEye, Earthstar Geographics, CNES/Airbus DS, USDA, USGS, AeroGrid, IGN, and the GIS user community

with 20–25 per site for the first 2 years, and 10 per site during the last 2 years. This sampling variation was accounted for in diversity analysis by extrapolating rarefaction curves to the same maximum (25 pans per site) using EstimateS version 9 (Colwell [2013](#page-11-14)). Down-sampling also assisted in accounting for differences in the number of sites at each grazing level. The pans were left undisturbed for 1 h of daylight, and then specimens were collected. All trapping was done during the daytime without storms, although control for uniform temperatures, weather conditions, and time of day was not possible. We sorted dipteran specimens to family; more specific taxon sorting could add resolution to our analysis, but was impractical given the number of specimens and time available for this analysis. We vouchered specimens from this study in the alcohol collection under Diptera in the M. L. Bean Life Science Museum at Brigham Young University. They are stored in 70% ethanol, in general with only one family per vial but perhaps more than one species in a vial.

### **Analysis**

We generated sample-based rarefaction curves using EstimateS (Colwell [2013\)](#page-11-14) to evaluate the effectiveness of our sampling at representing fly diversity for each level of grazing (Fig. [2](#page-6-0)). Species area/effort theory predicts that taxa will increase with effort up to a point of saturation of habitat, niche, or geographic footprint (Srivastava [1999](#page-12-6); Gotelli and Colwell [2001\)](#page-11-15), and we were interested in how close our efforts had come to saturation at each grazing level. The Est(S) function in EstimateS uses the reference sample to compute the expected number of different species in a given number of pooled samples (Colwell [2013\)](#page-11-14). Each sample

was represented by the number of flies or families of flies found in a single trap. We extrapolated each of the rarefaction curves to a total of 780 traps, corresponding to the total number of traps in our largest category (slight grazing) to account for uneven sampling. A grazing level was considered sampled to near-saturation if the number of observed families was more than 95% of what was expected after 780 traps. We randomized each curve with 100 sample order randomization runs with the EstimateS function, and compared the difference in rarefaction curves of family diversity at different grazing levels. If the 84% confidence intervals of the curves of different grazing levels did not overlap, a difference was considered significant (Fig. [3\)](#page-6-1). This metric mimics  $P < 0.05$  statistical testing more realistically than stricter confidence intervals (MacGregor-Fors and Payton [2013](#page-11-16)).

To analyze the effect of grazing intensity on terrestrial fly diversity, we compared fly abundance and family richness for each level of grazing using analysis of variance (ANOVA) in R version 3.2.2 (R Core Team [2013\)](#page-12-7). We also used EstimateS to compare diversity differences extrapolated for maximum or minimum real sampling effort (e.g., to 25 pans per site or with only 9 sites from each grazing level).

We evaluated how each family responded to different grazing pressures by grouping taxa into four categories based on the slope estimated from a linear regression model of that family's prevalance versus grazing intensity (Table [3](#page-7-0); Fig. [4](#page-8-0)). If the slope was less than  $-0.03$ , it was scored as having a negative response to grazing, being most prevalent at less-grazed sites (Fig. [4;](#page-8-0) Ceratopogonidae). If the slope was between −0.03 and 0.03 and the average frequency of the fly family was at least 10% higher in intermediatelygrazed areas (slight grazing, moderate grazing, or mixed grazing) than at the two extremes of grazing (no and heavy),

<span id="page-3-0"></span>Table 1 Collecting sites for yellow pan trap sampling, with grazing level (see Table [2](#page-5-0) for scale), number of pan traps set, and the number of fly families and individuals (both collected and as extrapolated to 25 pans)

Site name	Region	Locality	Graz- ing level $(0-4)$		Families Individuals Pans Est(S)		families at 25 pans	$Est(S)$ indi- viduals at 25 pans
2008_0201	Khovd	N48.01030° E91.61830° 1463 m	$\overline{c}$	8	60	25	8	60
2008_0202	Khovd	N48.33136° E92.81702° 1145 m	$\overline{c}$	14	279	23	14.39	303.26
2008_0301	Khovd	N48.32093° E91.30888° 1474 m	$\mathbf{1}$	9	114	20	9.61	142.5
2008_0302	Khovd	N48.42161° E90.97337° 1805 m	$\overline{c}$	19	197	25	19	197
2008_0401	Bayan-Ölgii	N48.42305° E90.38785° 2310 m	$\sqrt{2}$	7	7	25	7	7
2008_0403	Bayan-Ölgii	N48.64831° E89.88409° 2066 m	$\mathbf{1}$	9	29	21	9.21	34.52
2008_0501	Bayan-Ölgii	N48.97842° E89.94995° 1750 m	4	7	23	22	7.52	26.14
2008_0502	Bayan-Ölgii	N48.64815° E89.88378° 2065 m	$\mathbf{1}$	12	224	21	12.3	266.67
2008_0601	Bayan-Ölgii	N48.83461° E89.19676° 2055 m	$\overline{c}$	15	318	21	15.34	378.57
2008_0602	Bayan-Ölgii	N49.04180° E89.41676° 1780 m	2	$18\,$	335	22	18.5	380.68
2008_0603	Bayan-Ölgii	N49.12999° E89.37314° 1798 m	$\mathbf{1}$	15	238	22	15.59	270.45
2008_0701	Bayan-Ölgii	N49.23798° E88.90710° 2104 m	$\mathbf{1}$	11	154	22	11.1	175
2008_0702	Bayan-Ölgii	N49.23477° E88.90018° 2101 m	$\mathbf{1}$	11	113	20	11.93	141.25
2008_0801	Bayan-Ölgii	N49.38138° E88.6839° 2304 m	$\mathbf{1}$	18	343	22	18.19	389.77
2008_0802	Bayan-Ölgii	N49.31761° E88.37029° 2382 m	$\mathbf{1}$	11	250	22	11.51	278.41
2008_1002	Bayan-Ölgii	N49.12007° E88.01157° 2912 m	$\mathbf{1}$	18	445	21	18.98	529.76
2008_1003	Bayan-Ölgii	N49.15035° E87.94238° 3099 m	$\mathbf{1}$	10	685	21	10.9	815.48
2008_1302	Bayan-Ölgii	N49.04586° E88.50939° 2385 m	$\mathbf{1}$	19	671	21	19.53	798.81
2008_1304	Bayan-Ölgii	N48.75576° E88.60196° 2145 m	$\mathbf{1}$	16	128	22	17.06	145.45
2008_1305	Bayan-Ölgii	N48.76028° E88.60029° 2176 m	$\mathbf{1}$	16	182	22	16.31	206.82
2008_1401	Bayan-Ölgii	N48.72628° E88.40073° 2439 m	3	15	133	22	15.77	151.14
2008_1402	Bayan-Ölgii	N48.66718° E88.29908° 2090 m	3	7	299	21	7.3	355.95
2008_1501	Bayan-Ölgii	N48.75577° E88.15264° 2115 m	$\mathbf{1}$	$18\,$	177	21	18.39	210.71
2008_1502	Bayan-Ölgii	N48.75197° E88.14987° 2098 m	$\mathbf{1}$	13	120	22	13.75	136.36
2008_1503	Bayan-Ölgii	N48.54381° E88.41416° 2147 m	$\overline{c}$	12	212	21	12.68	252.38
2008_1601	Bayan-Ölgii	N48.62918° E88.28554° 2147 m	$\mathbf{1}$	13	384	20	14.86	481.25
2008_1604	Bayan-Ölgii	N48.50679° E88.50977° 2129 m	$\mathbf{1}$	15	233	22	15.36	264.77
2008_1703	Bayan-Ölgii	N48.16728° E88.85435° 2073 m	$\overline{c}$	19	253	22	20.04	287.5
2008_1803	Bayan-Ölgii	N48.34519° E89.32607° 2438 m	$\mathbf{1}$	13	85	21	13.86	101.19
2008_1901	Bayan-Ölgii	N48.24120° E89.60174° 2150 m	$\sqrt{2}$	17	233	21	18.01	277.38
2008_1902	Bayan-Ölgii	N48.54136° E89.55995° 2029 m	$\mathbf{1}$	13	56	21	14.29	66.67
2008_2001	Khovd	N48.41182° E90.891° 1922 m	3	13	230	22	13.36	261.36
2008_2002	Khovd	N47.94771° E91.55796° 1447 m	$\overline{c}$	12	124	21	13.7	147.62
2009_0101	Khovd	N47.97652° E91.59132° 1428 m	3	12	94	25	12	94
2009_0401	Khovd	N47.53183° E091.28320° 2519 m 2		$10\,$	68	$25\,$	10	68
2009_0402	Khovd	N47.53169° E091.28284° 2521 m 2		9	29	25	9	29
2009_0403	Bayan-Ölgii	N47.58121° E091.17047° 2049 m 2		12	89	25	12	89
2009_0405	Bayan-Ölgii	N47.84396° E090.64262° 2165 m 2		8	36	25	8	36
2009_0501	Bayan-Ölgii	N47.62447° E090.67194° 2241 m 3		22	262	25	22	262
2009_0502	Bayan-Ölgii	N47.66395° E090.71841° 2196 m 3		10	166	25	10	166
2009_0601	Bayan-Ölgii	N47.45456° E090.92342° 2631 m 2		6	61	25	6	61
2009_0602	Bayan-Ölgii	N47.34660° E090.96013° 2519 m 2		9	93	25	9	93
2009_0604	Bayan-Ölgii	N47.08868° E091.02686° 2056 m 2		13	88	25	13	$88\,$
2009_0704	Bayan-Ölgii	N47.24368° E090.75253° 2560 m 2		14	122	25	14	122
2009_0705	Bayan-Ölgii	N47.18333° E090.77660° 2492 m 3		18	985	$25\,$	18	985
2009_0802	Bayan-Ölgii	N47.03794° E091.02931° 2016 m 3		12	153	25	$12\,$	153
2009_0803	Bayan-Ölgii	N47.03954° E091.03448° 2010 m 3		9	63	25	9	63



**Table 1** (continued)



<span id="page-5-0"></span>



we categorized it as having an intermediate response, being most abundant at middle amounts of grazing (Fig. [4;](#page-8-0) Anthomyiidae). If the slope was greater than 0.03, the family was considered to have a positive response to grazing pressures, being more prevalent at heavily grazed areas (Fig. [4;](#page-8-0) Ephydridae). Finally, a family with a slope between −0.03 and 0.03 and with prevalence at middle grazing pressures less than 10% higher than at extremes was categorized as having

<span id="page-6-0"></span>

<span id="page-6-1"></span>**Fig. 3** Diptera family richness for different grazing intensities with an 84% confidence interval as a response to effort at 90 pans (number at least-sampled grazing level), 300 pans, 500 pans and 780 pans

(number at most-sampled grazing level). Diversity metrics have been extrapolated to show expectations under a uniform sampling effort using EstimateS

no response (Fig. [4;](#page-8-0) Phoridae). Data-deficient taxa (a family with fewer than 10 individuals collected) were not categorized as their rarity would not allow for robust statistical testing. The thresholds for slope categories were set at −0.03 and 0.03 because families with slopes >  $0.03$  or  $<-0.03$  also had correlation coefficients >  $0.5$  and  $<-0.5$  respectively.

# **Results**

We visited 132 different sites and set up 2334 pan traps to collect 17,348 flies. This yielded, on average, 7.0 flies per trap (ranging from 0 to 115) and 2.6 different families in each trap. Of the 155 recognized families of Diptera (Marshall [2012\)](#page-11-17), 49 were represented in our samples (Table [3](#page-7-0)). A third of the individuals we collected were Chironomidae (5858 from 112 sites). Other prominent families were Ephydridae (2459 from 112 sites), Dolichopodidae (2273 from 105 sites), Muscidae (1922 from 105 sites), Anthomyiidae (723 from 86 sites), Chloropidae (688 from 90 sites), Phoridae (587 from 53 sites), Tachinidae (309 from 58 sites) and Sciaridae (303 from 76 sites). In all, 87% of the flies we collected were from these nine families. Six families were collected as a single individual: Aulacigastridae, Conopidae, Lonchopteridae, Opomyzidae, Sciomyzidae, and Therevidae. Most of the 49 families we collected had intermediate levels of abundance between 10 and 300 individuals. The families that were encountered at the highest frequencies also had the greatest abundance (Fig. [5\)](#page-8-1).

<span id="page-7-0"></span>**Table 3** Number and percent of sites where each of the families was found at each grazing level

Family	No grazing (9 sites)	Slight grazing $(48$ sites)	Moderate grazing $(33 \text{ sites})$	Mixed grazing $(27$ sites)	Heavy grazing $(11$ sites)	Slope	Response to grazing
Agromyzidae	2(22%)	11 (23%)	7(21%)	9(33%)	3(27%)	0.02	None
Anthomyiidae	3(33%)	32 (67%)	28 (85%)	20 (74%)	2(18%)	$-0.02$	Intermediate
Asilidae	1(11%)	2(4%)	2(6%)	3(11%)	$0(0\%)$	$-0.02$	None
Aulacigastridae	$0(0\%)$	$0(0\%)$	$0(0\%)$	$0(0\%)$	1(9%)	0.02	DD
Bibionidae	$0(0\%)$	$0(0\%)$	2(6%)	$0(0\%)$	$0(0\%)$	$\boldsymbol{0}$	DD
Blephariceridae	$0(0\%)$	$5(10\%)$	1(3%)	1(4%)	$0(0\%)$	$-0.01$	None
Bombyliidae	1(11%)	4(8%)	6(18%)	1(4%)	2(18%)	0.01	None
Calliphoridae	1(11%)	12(25%)	11 (33%)	11 $(41%)$	$0(0\%)$	$-0.01$	Intermediate
Carnidae	$0(0\%)$	$5(10\%)$	5(15%)	7(26%)	$0(0\%)$	0.02	Intermediate
Cecidomyiidae	$0(0\%)$	12(25%)	11(33%)	6(22%)	$0(0\%)$	$\boldsymbol{0}$	Intermediate
Ceratopogonidae	5(56%)	24 (50%)	16 (48%)	8 (30%)	2(18%)	$-0.10$	Negative
Chamaemyiidae	$0(0\%)$	$0(0\%)$	1(3%)	2(7%)	$0(0\%)$	0.01	None
Chironomidae	8 (89%)	40 (83%)	30 (91%)	22 (81%)	8 (73%)	$-0.03$	Negative
Chloropidae	4(44%)	29 (60%)	25 (76%)	22 (81%)	7(64%)	0.06	Positive
Conopidae	$0(0\%)$	$0(0\%)$	1(3%)	$0(0\%)$	$0(0\%)$	$\boldsymbol{0}$	DD
Culicidae	$0(0\%)$	6(13%)	2(6%)	1(4%)	$0(0\%)$	$-0.01$	None
Dolichopodidae	6(67%)	42 (88%)	27 (82%)	24 (89%)	6(55%)	$-0.02$	Intermediate
Drosophilidae	$0(0\%)$	1(2%)	1(3%)	2(7%)	$0(0\%)$	0.01	DD
Empididae	$1(11\%)$	21 (44%)	16 (48%)	15 (56%)	2(18%)	0.03	Intermediate
Ephydridae	6(67%)	43 (90%)	25 (76%)	23 (85%)	11 (100%)	0.06	Positive
Heleomyzidae	$0\ (0\%)$	2(4%)	1(3%)	2(7%)	$0(0\%)$	$\boldsymbol{0}$	DD
Lauxaniidae	$1(11\%)$	$0(0\%)$	$0(0\%)$	1(4%)	$0(0\%)$	$-0.02$	DD
Lonchopteridae	$0(0\%)$	$0(0\%)$	1(3%)	$0(0\%)$	$0(0\%)$	$\boldsymbol{0}$	DD
Micropezidae	$1(11\%)$	3(6%)	3(9%)	8 (30%)	$0(0\%)$	$\boldsymbol{0}$	None
Muscidae	5(56%)	40 (83%)	25 (76%)	22 (81%)	$10(91\%)$	0.07	Positive
Mycetophilidae	$0(0\%)$	2(4%)	1(3%)	$0(0\%)$	$0(0\%)$	$\boldsymbol{0}$	DD
Opomyzidae	$1(11\%)$	$0(0\%)$	$0(0\%)$	$0(0\%)$	$0(0\%)$	$-0.02$	DD
Phoridae	3(33%)	22 (46%)	17 (52%)	7(26%)	4(36%)	$-0.01$	None
Pipunculidae	$0(0\%)$	2(4%)	$0(0\%)$	$0(0\%)$	1(9%)	0.01	DD
Psychodidae	$1(11\%)$	$5(10\%)$	4(12%)	4(15%)	2(18%)	0.02	None
Rhagionidae	$0(0\%)$	1(2%)	$0(0\%)$	1(4%)	$0(0\%)$	$\boldsymbol{0}$	DD
Sarcophagidae	4(44%)	14 (29%)	12 (36%)	10(37%)	3(27%)	$-0.03$	Negative
Scathophagidae	$0(0\%)$	3(6%)	2(6%)	3(11%)	$0(0\%)$	$\boldsymbol{0}$	None
Scatopsidae	$0(0\%)$	$5(10\%)$	2(6%)	6(22%)	$0(0\%)$	0.01	Intermediate
Sciaridae	7(78%)	27 (56%)	20 (61%)	14 (52%)	6(55%)	$-0.05$	Negative
Sciomyzidae	$0\ (0\%)$	1(2%)	$0(0\%)$	$0(0\%)$	$0\ (0\%)$	$\boldsymbol{0}$	DD
Sepsidae	2(22%)	13 (27%)	12 (36%)	10 (37%)	4(36%)	0.04	Positive
Simuliidae	1(11%)	6(13%)	8(24%)	2(7%)	1(9%)	$-0.01$	None
Sphaeroceridae	3(33%)	19 (40%)	14 (42%)	14 (52%)	4(36%)	$0.02\,$	Intermediate
Stratiomyidae	$0(0\%)$	4(8%)	3(9%)	4(15%)	1(9%)	0.02	None
Syrphidae	2(22%)	9(19%)	3(9%)	7(26%)	2(18%)	$\boldsymbol{0}$	None
Tabanidae	$0(0\%)$	2(4%)	1(3%)	$0(0\%)$	1(9%)	0.01	$\rm{DD}$
Tachinidae	4 (44%)	18 (38%)	18 (55%)	12(44%)	6(55%)	0.03	Positive
Tanypezidae	$1(11\%)$	$0(0\%)$	1(3%)	1(4%)	$0(0\%)$	$-0.02$	DD
Tephritidae	$0(0\%)$	1(2%)	2(6%)	2(7%)	$0(0\%)$	$0.01\,$	DD
Therevidae	1(11%)	$0(0\%)$	$0(0\%)$	1(4%)	$0(0\%)$	$-0.02$	DD
Tipulidae		16 (33%)	7(21%)			$-0.02$	None
	2(22%)			7(26%)	2(18%)		
Trichoceridae Ulidiidae	$0(0\%)$ $0(0\%)$	1(2%) $5(10\%)$	$0(0\%)$ $0(0\%)$	$0(0\%)$ 4(15%)	$0(0\%)$ $0(0\%)$	$\boldsymbol{0}$ $\boldsymbol{0}$	DD None

Response categories are based on slope and frequency differences between extreme and intermediate levels

*DD* data deficient families with fewer than 10 occurrences



<span id="page-8-0"></span>**Fig. 4** Example of differential responses of select Diptera families to grazing pressure as gathered from yellow pan traps in Mongolia. Ceratopogonidae displays a negative response to increased grazing pressure. Anthomyiidae is most prevalent in areas of intermediate dis-

turbance. Ephydridae shows a positive response to increased grazing pressures, and Phoridae has a negligible response to grazing. Dotted lines represent line of best fit. Solid lines connect values from different grazing pressures

### **Sampling effectiveness**

The richness accumulation curves showed that we collected enough samples to reach near-saturation values of 95% or more of the total extrapolated diversity for each grazing level except no grazing, where we collected only 63% of the diversity (Fig. [2\)](#page-6-0). Our sampling yielded 26 diferent families from ungrazed sites (from 90 pans), 40 different families from sites with slight grazing (780 pans), 40 families from sites with moderate grazing (636 pans), 36 families from sites with mixed grazing (508 pans) and 24 families from sites with heavy grazing (122 pans).



<span id="page-8-1"></span>**Fig. 5 a** Rank abundance plot of the total number of flies from each family collected at all sites and **b** frequency of each family at 132 sites ordered by abundance

#### **Richness and abundance across a grazing gradient**

The overlapping 84% confidence intervals of our extrapolated species effort curves (Fig. [3](#page-6-1)) indicate that heavily grazed sites have lower overall fly family diversity than sites with intermediate levels of grazing (e.g., slight, moderate, and mixed). The confidence interval for ungrazed sites remained extremely broad (likely due to the small true sample size for this category), and overlapped with the confidence interval for heavily grazed sites. These relationships remained consistent at both our simulated high and low sampling schemes (e.g., 780 pans, the maximum at any category, and 90 pans, the minimum at any category). Similarly, without extrapolation, mean numbers of fly families at sites with intermediate grazing levels (slight: 8.88, moderate: 8.85, mixed: 8.96) are higher than the average number of flies at heavy grazing (8.18) and no grazing (8.44), however, these were not statistically significant (ANOVA,  $F = .2218$ ,  $df = 4$ ,  $p = 0.9258$ ).

Average overall Diptera abundance was greater at sites with grazing than at sites with no grazing (Fig. [6](#page-9-0)), but these differences were not statistically significant (ANOVA,  $F=0.349$ , df = 4, p = 0.8443).

### **Taxonomic response to grazing**

Different fly families spanned the variety of possible responses to livestock grazing pressure (Table [3](#page-7-0); Fig. [4](#page-8-0)). Four families (Chironomidae, Sciaridae, Ceratopogonidae and Sarcophagidae) decreased in frequency as grazing increased, eight families (Dolichopodidae, Anthomyiidae,



<span id="page-9-0"></span>**Fig. 6** Abundance of diptera at different grazing pressures using the Est(s) number of individuals found at each site extrapolated to 25 pans per site

Empididae, Sphaeroceridae, Cecidomyiidae, Calliphoridae, Carnidae, and Scatopsidae) were most prevalent at intermediate grazing, and five families (Ephydridae, Muscidae, Chloropidae, Tachinidae and Sepsidae) increased in frequency as grazing increased. In addition, 32 families had no responses to grazing or did not have enough specimens collected to make conclusive judgements about patterns. However, in the 23 families where 20 or fewer specimens were collected, only three of them, Pipunculidae, Tabanidae, and Aulacigastridae, were collected in areas with heavy grazing. Of the nine most abundant families, all but Phoridae showed a positive, negative, or intermediate response.

# **Discussion**

Our project represents the most complete effort thus far to document flies in Mongolia. In our 17,348 flies, we encountered 49 different families. This is comparable to the results of similar surveys done in Australian rain forests where 56 families were found from 28,647 flies (Kitching et al. [2004\)](#page-11-18) or in a forested national park in Germany where 62 families were found among 19,641 individuals (Scherber et al. [2014](#page-12-8)).

An overall aim of the project was to collect fly families at every site that had been chosen by the MAIS survey. We had no expectation of sampling lightly and heavily sites equally, and in the end had more moderately grazed sites than ungrazed or heavily grazed sites, so it was necessary to rarify our data for comparison. This analysis shows that fly diversity at our sites in Mongolia is greater at intermediate levels of grazing (slight, moderate, mixed) than in heavily grazed areas. This may be consistent with Connell's intermediate-disturbance hypothesis [\(1978\)](#page-11-4) or with a decrease in diversity with increased grazing. At sites with heavy grazing, there is less variation in family diversity and fewer families than at sites with intermediate levels of grazing. However, streams where no grazing was observed have a high variation in fly family diversity that is sometimes as low as the heavily grazed sites. More ungrazed streambanks need to be sampled to better resolve the impact of grazing in this environment, though locating such sites may be difficult as commercialized herding spreads across the country.

In contrast to the decrease in family diversity, Diptera abundance increased in sites where at least some grazing was present. Part of this rise was due to immense abundance increases for specific families at single sites. For example, in the family Phoridae, 587 individuals were collected throughout the course of the study, of which 324 were collected from a single heavily grazed site. This is one of the most ecologically diverse groups of flies, and it is not unheard of for this family to dominate the abundance at a particular site (Brown [2005\)](#page-11-19). At another site that had mixed grazing, 828 Chironomidae individuals were collected. Additionally, 209

of the 243 Psychodidae individuals were also collected from a single site with mixed grazing. Both of these families have members that are especially well-adapted to anoxic environments as larvae (Boltz et al. [2008](#page-11-20)), which may explain their high abundance in areas where grazing mammals provide the necessary habitat in a localized manner (Verdú et al. [2000\)](#page-12-1). Such single-site superabundances can skew observed trends, so it is important to be mindful of the influence of outliers on parametric statistics when studying highly heterogeneous ecological systems.

Four of the five families that increased in abundance with grazing pressures, Ephydridae, Muscidae, Tachinidae and Sepsidae, are commonly found in connection with animal biproducts such as carrion or animal excrement (McAlpine et al. [1981](#page-11-5)). The fifth, Chloropidae, is mainly a plant feeder, but includes many saprophagous species which often suck up liquid secretions and tend to mass around animals, including at least one genus that feeds in excrement or dead animals (Oldroyd [1965;](#page-11-21) Ismay and Nartshuk [2000\)](#page-11-22). As more livestock are grazed, greater amounts of animal excrement provide habitat for these families. However, many other dung-feeding or dung-breeding families like Calliphoridae, Sphaeroceridae, Scatopsidae, Phoridae, Psychodidae, Carnidae and Sarcophagidae did not increase with increased grazing. In fact, Calliphoridae was not collected at any sites with heavy grazing, and Sarcophagidae responded negatively to increased grazing. Following a disturbance, insects that are more tolerant to stress may dominate a community (Schowalter [2012\)](#page-12-9), which may explain why some taxa ostensibly adapted for the byproducts of grazing nevertheless found themselves shunted into a minor role.

The families that responded negatively to increased grazing pressure (Chironomidae, Sciaridae, Sarcophagidae, and Ceratopogonidae) are found in a variety of habitats. Most Chironomidae are aquatic as larvae, but as terrestrial adults live on a variety of substrates, Sciaridae are often vegetable feeders or live on fungi, Sarcophagidae feed on flesh or carrion, and Ceratopogonidae feed on either other insects or mammals (Oldroyd [1965](#page-11-21); McAlpine et al. [1981](#page-11-5)). Each of these families have members that inhabit a wide variety of niches but all of these habitats could be degraded or impacted with increased grazing pressures.

In aquatic assessment protocols, Chironomidae have historically been used as indicators of high levels of disturbance or stress (Pinder [1986](#page-12-10); Plafkin et al. [1989](#page-12-11)), often the dominant macroinvertebrate in heavily polluted habitats (Winner et al. [1980\)](#page-12-12). However, the group is taxonomically diverse and Chironomidae species can be found in almost every body of freshwater (Pinder [1986](#page-12-10)), so characterizing all of Chironomidae as "tolerant" of pollution or disturbances is an over-generalization. Although chironomids were prevalent in high numbers at a majority of our sites, they showed the greatest affinity for sites where there was low or intermediate amounts of grazing. We suspect that the rich abundance and diversity of aquatic forms of chironomids at our sites may swamp any signal from the less diverse soil inhabiting guild of these flies that has occasionally been reported in previous ecological surveys (Frouz [1999\)](#page-11-9).

The family where we observed the most dramatic negative response to livestock grazing is Ceratopogonidae, the "biting midges". Diptera from this family were present at more than 50% of ungrazed or slightly grazed sites, while we encountered them only twice in our 18 heavily grazed sites. Members of this family have been found to be responsive to pesticides in other habitats (Rochlin et al. [2011\)](#page-12-13) and this may reflect their susceptibility to other environmental disturbances such as grazing, and potential as bioindicators of acceptable levels of grazing.

We were surprised by a lack of response in several families where a response was expected. For example, aquatic crane flies in Mongolia responded negatively to grazing pressures in a previous stucdy (Yadamsuren et al. [2015](#page-12-0)), but we encountered them at similar frequencies across all levels of grazing. This may be because the larvae respond differently than the adults or because this effect can only be seen at the species level.

The families with the most visible responses to grazing were Ceratopogonidae as an indicator of low grazing pressures and Ephydridae and Muscidae as indicators of heavy grazing. We recommend these families as indicators of grazing pressures. Future research should focus on further evaluating these families as bioindicators in areas with similar or distinct biota. These families could also be the focus of future work at the species level.

In conclusion, the consequences of large livestock mammal grazing on fly populations in Mongolia are complex and tailored to individual taxonomic groups, but overall, heavy grazing has a negative effect on fly diversity at the family level. The same pattern is likely reflected at the species level, but we need more intensive field experiments to begin to speculate the reasons why. Although many fly families have members that are part of several different guilds and it may be easier to measure guilds within families, differences in habitat preferences have been documented in guilds of fly families (Hughes et al. [2000](#page-11-23)), and we can begin to see the makeup of these guilds in the results of our study. As levels of commercial grazing continue to increase in Mongolia, many populations of flies, especially those dependent on low levels of grazing are vulnerable. Understanding fly diversity metrics and taxonomic guilds is important to understanding the effect that grazing disturbances have on biodiversity as a whole and may be useful for evaluating land use.

**Acknowledgements** We thank directing members of the Mongolian Aquatic Insect Survey, listed below, for their support in this project. We also thank the participants and support staff of the Mongolian Aquatic

Insect Survey. We thank members of the Nelson Laboratory at Brigham Young University, especially Joss Wertz, who collected the first year of data for this project. We thank the Roger and Victoria Sant Endowment for a Sustainable Environment (Brigham Young University) for providing funding for this project to get students into the unfamiliar territory of central Asia. We gratefully acknowledge the generous support of the US National Science Foundation (DEB-BSI #0743732, "Survey and Inventory of the Aquatic Macroinvertebrates of the Altai and Hangai Mountain drainages, Mongolia", to J. Gelhaus, J. Morse, C. R. Nelson) for direct support of this fly diversity research and to (DEB-BSI #0206674, "Survey and Inventory of the Aquatic Macroinvertebrates of the Selenge River, Mongolia" to J. Gelhaus, J. Morse, B. Hayford), and (DEB-BS&I #0816910, "An Ecological Guild-Based Biodiversity Inventory and Survey of the Aquatic Non-biting Midges (Diptera: Chironomidae) of the Altai and Hangai Mountain Drainages, Mongolia" to B. Hayford) for supporting work. Our studies were conducted under the arching support of the Academy of Natural Sciences of Drexel University, Clemson University, Brigham Young University, and Wayne State College. We give special thanks to Jon Gelhaus, Barbara Hayford, the late Sarah Walker Judson, Jigmed Khandsuren "Handaa", Alain Maasri, John Morse, Sigitas Podenas, Virginija Podeniene, Chuluunbat Suvdtsetseg, Sundarea, Enkhnasan Davaadorj for their field help and the entire MAIS team for their hard work and diligence. We give our highest thanks and respect to the late Balgan Namkhaidorj for his

professional entomological guidance, his uncanny geographical knowledge, and his direct material safety mentoring during our expeditions as we bounced across the steppe and taiga. Goodbye friend, bayarlalaa. Any opinions, findings, and conclusions or recommendations expressed in this material are those of the authors and do not necessarily reflect the views of the US National Science Foundation.

### **Compliance with ethical standards**

**Conflict of interest** The authors declare that they have no conflict of interest.

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